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Review

Zwitterionic glycosidase inhibitors: salacinol and related analogues Sankar Mohan and B. Mario Pinto*

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Abstract—Natural products with interesting biological properties and structural diversity have often served as valuable lead drug candidates for the treatment of human diseases. Salacinol, a naturally occurring α -glucosidase inhibitor, was shown to be one of the active principles of the aqueous extract of a medicinal plant that has been prescribed traditionally as an Ayurvedic treatment for type II diabetes. Salacinol contains an intriguing zwitterionic sulfonium-sulfate structure that comprises a 1,4-anhydro-4-thio-p-arabinitol core and a polyhydroxylated acyclic chain. Due to the unique structural features and its potential to become a lead drug candidate in the treatment of type II diabetes, a great deal of attention has been focused on salacinol and its analogues. Since the isolation of salacinol, several papers describing various synthetic routes to salacinol and its analogues have appeared in the literature. This review is aimed at highlighting the synthetic aspects of salacinol and related compounds as well as their structure–activity relationship studies.

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1. Introduction

Over the years, glycosidase inhibitors have received considerable attention in the field of chemical and medicinal research. They have many potential therapeutic applications because the glycosidase enzyme-catalyzed hydrolysis of complex carbohydrates is a biologically widespread phenomenon in living systems.^{2–4} For example, inhibition of glycosidase enzymes that are involved in the biosynthesis of oligosaccharide chains of the Nlinked glycoproteins in the endoplasmic reticulum (ER) and Golgi apparatus has profound effects on maturation, transport, and secretion of these glycoproteins.^{5,6} This strategy has potential for many therapeutic applications, such as in the treatment of cancer and viral infections.⁵ Inhibition of carbohydrate-hydrolyzing enzymes such as pancreatic α -amylase and α -glucosidases in the digestive tract that leads to a delay in digestion of ingested carbohydrates is one of the therapeutic approaches for the treatment of type II non-insulin-dependent diabetes mellitus (NIDDM).^{7,8}

Many naturally occurring monocyclic and bicyclic amines such as 1-deoxynojirimycin (1), swainsonine (2), and castanospermine (3) (Fig. 1) are effective inhibitors of various glycosidase enzymes and have shown potential as therapeutic agents. For example, treatment with the indolizidine alkaloid swainsonine (2), a naturally occurring Golgi α -mannosidase II inhibitor, has led to significant reduction of tumor mass in human patients with advanced malignancies, and is a promising drug therapy for patients suffering from breast, liver, lung, and other malignancies. 10,11

These amine-based glycosidase inhibitors are believed to carry a positive charge at physiological pH and hence are postulated to bind in the active sites of glycosidase enzymes by mimicry of the charge of the oxacarbenium-ion-like transition state formed during hydrolysis of the natural substrate (Fig. 2).¹² The establishment of stabilizing electrostatic interactions between the protonated nitrogen and an active-site carboxylate residue is

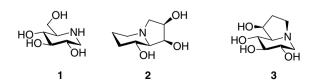


Figure 1. Examples of naturally occurring glycosidase inhibitors: 1-deoxynojirimycin (1), swainsonine (2), and castanospermine (3).

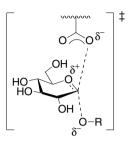


Figure 2. Oxacarbenium-ion-like transition state for the glycosidase-mediated hydrolysis reaction.

considered to be a possible mechanism of action of such inhibitors. 12

Hence, while designing a synthetic inhibitor, consideration of inclusion of an atom that carries a permanent positive charge at a suitable position was considered as an alternative means of providing the required charge state to establish such electrostatic interactions between the inhibitor and the active-site carboxylate residue. Accordingly, introduction of a permanent positive charge on the nitrogen atoms of azasugars was attempted. For example, Kajimoto et al. ¹³ synthesized the *N*-oxide analogue of castanospermine (4) shown in Figure 3. However, this compound was only a weak inhibitor of β-glucosidase compared to the parent compound, castanospermine (3).

For the purpose of establishing such electrostatic pairing, introduction of a positively charged sulfur atom in the structures of potential glycosidase inhibitors was considered next. The approach had precedent in the mimicry by sulfonium ions of tertiary amines with narcotic analgesic properties. For example, Belleau et al. 14-17 reported the syntheses of sulfonium-ion analogues of levorphanol (5) and isolevorphanol (6), Smethyl sulforphanol (7), and S-methyl isosulforphanol (8) (Fig. 3), respectively, and showed that they were agonists or antagonists of morphine for the opiate receptor. In addition, the syntheses of various sulfonium salts such as the bicyclic compounds 9¹⁸ and 10, ¹⁹ and a sulfimide derivative 11²⁰ (Fig. 3) have been reported, and these compounds have been tested for their glycosidase inhibitory activities. We have reported the synthesis of a sulfonium-ion analogue of castanospermine, 12 (Fig. 3), as a model to test the theory that a sulfonium salt carrying a permanent positive charge might be advantageous in providing the necessary electrostatic interactions between the inhibitor and the active-site carboxylate residues.²¹ After initiation of this work, a

Figure 3. Synthetic glycosidase inhibitors carrying a permanent positive charge.

Figure 4. Structures of salacinol (13) and kotalanol (14).

literature report strongly validated this approach as two naturally occurring glycosidase inhibitors, namely salacinol (13)²² and kotalanol (14),²³ both having a sulfonium-ion center (Fig. 4), were isolated from *Salacia reticulata*, known as 'Kotalahimbutu' in Singhalese, a large woody climbing plant widespread in Sri Lanka and South India. Aqueous extracts of this plant have been traditionally used in the Ayurvedic system of Indian medicine for treating type II diabetes.^{22–24} Traditionally, Ayurvedic medicine advised that a person suffering from diabetes should drink water stored overnight in a mug carved from Kotalahimbutu wood.

It was demonstrated that these two natural products were active components of the aqueous extracts of this plant. Salacinol (13) and kotalanol (14) share a common zwitterionic sulfonium-sulfate structure that comprises a 1,4-anhydro-4-thio-p-arabinitol core and a polyhydroxylated acyclic chain, the difference between them being the length of the polyhydroxylated acyclic chain; kotalanol has a seven-carbon chain, with unspecified (to date) stereochemistry at the stereogenic centers, whereas salacinol has a four-carbon chain with defined stereochemistry. Although the exact mechanism of action of salacinol and kotalanol has yet to be elucidated, it is postulated that the positively charged sulfur atom can act in the same way as the protonated ammonium center of the amine-based glycosidase inhibitors when binding in the active site of glucosidases that process carbohydrate polymers and oligomers. 25,26

2. Salacinol and its analogues

2.1. Isolation and structural elucidation of salacinol

Salacinol (13) was originally isolated from S. reticulata by Yoshikawa et al.²² and later also from other plant species of the Salacia genus, Salacia oblonga²⁷ and Salacia chinensis, 28 by the same group. Through bioassayguided separation using \alpha-glucosidase inhibitory activities, a potent glycosidase inhibitor that they named salacinol (13) was isolated from the water-soluble fractions obtained from the dried roots of S. reticulata.²² The water-soluble fraction inhibited the increase in serum glucose levels after the administration of sucrose or maltose in rats. Column chromatographic purification of the water-soluble fraction followed by repeated HPLC separations gave the pure natural product salacinol (13). The purified salacinol showed competitive inhibition for intestinal α-glucosidases in vitro: the glucosidase inhibitory activities of salacinol against maltase and sucrase were nearly equal to those of acarbose, a clinically used α -glucosidase inhibitor of microbial origin (Table 1). However, the inhibitory activity of salacinol against isomaltase was found to be greater than that of acarbose.

The structural elucidation studies of salacinol used various chemical and spectroscopic methods including 1D and 2D NMR experiments and a potassium rhodizonate test (a positive test that confirmed the presence of a sulfate moiety), and suggested the presence of an

Table 1. K_i values of salacinol (13) for rat small-intestinal disaccharidases²²

Enzyme	$K_{\rm m}$ (M)	$K_{\rm i}~(\mu {\rm g/mL})$					
		Salacinol (13)	Acarbose				
Maltase	2.7×10^{-3}	0.31	0.12				
Sucrase	2.0×10^{-2}	0.32	0.37				
Isomaltase	4.5×10^{-3}	0.47	75				

Figure 5. Structure of salacinol as initially proposed by Yoshikawa et al.²²

unusual zwitterionic sulfonium-sulfate structure. Based on the spectral and X-ray crystallographic analyses of salacinol (13), Yoshikawa et al.²² assigned the initial stereostructure of salacinol as an inner-salt sulfonium structure (15) that was composed of a 1,4-anhydro-4-thio-L-arabinitol unit alkylated at sulfur by a 1-deoxy-D-erythritol-3-sulfate unit, as shown in Figure 5.

However, the degradation studies of kotalanol (14), also performed by Yoshikawa et al., ²³ led to the revision of the initial stereostructure (15) of salacinol. In this study, although the configurations of the stereogenic centers in the longer heptitol side chain of kotalanol (14) or at the sulfur atom were not determined, the presence of an anhydro-4-thio-D-arabinitol unit was con-

firmed based on the degradation product, 1,4-anhydro-4-thio-p-arabinitol (16), that was formed upon alkaline treatment of kotalanol (14) (Scheme 1). Because salacinol (13) and kotalanol (14) are of the same plant origin and are likely to arise from similar biosynthetic pathways, the structure of salacinol was then assigned to be the enantiomer (13) of the initial stereostructure (15).^{27,29}

2.2. Synthesis of salacinol

The ambiguity in the structure of salacinol was resolved when Yuasa et al.³⁰ and we³¹ independently reported the total synthesis of salacinol (13). In 2000, Yuasa et al.³⁰ first reported the synthesis of salacinol (13) and its diastereomer 17 (Scheme 2). The key step used for the synthesis of salacinol, involved a ring-opening reaction of the cyclic sulfate 18, derived from L-glucose, with 1,4-anhydro-4-thio-D-arabinitol (16),²⁰ followed by removal of the protecting groups (Scheme 2). This study indicated that the unprotected thioether 16 reacted smoothly only with the cyclic sulfates 18 and 19, that are protected as an isopropylidene ketal, when compared to other cyclic sulfates protected with benzyl or

Scheme 1.

benzylidene groups, or without protecting groups; in the latter cases, decomposition of the cyclic sulfates was observed.

We achieved the synthesis of salacinol (13) and its stereoisomers 15 and 17 via nucleophilic ring opening of a benzylidene-protected cyclic sulfate (20, derived from L-glucose or 21, derived from D-glucose), similar to the approach of Yuasa et al., 30 but using a per-benzylated thioether 22³² or 23³³ (Scheme 3). 31 Finally, the protecting groups were removed by hydrogenolysis. In this synthesis, dry acetone was found to be a more suitable solvent for the coupling reaction and the addition of K₂CO₃ in the reaction mixture was necessary to pre-

vent the unwanted hydrolysis reaction of the cyclic sulfate.

A significant improvement in this synthesis was achieved by exploiting an unusual solvent effect provided by 1,1,1,3,3,3-hexafluoroisopropanol (HFIP).³⁴ Thus, the reaction of the per-benzylated thioether **22** and the cyclic sulfate **20** in HFIP containing K₂CO₃ gave a significantly increased yield of the alkylated product (94%). The increased yields in HFIP were considered to be the effect of better solvation of the transition states relative to the ground states of the reactants. However, a relatively low yield of the final product (65%) was observed in the hydrogenolysis step (Scheme 4a), likely

Scheme 3.

Scheme 4b.

due to the poisoning of the catalyst by the small amount of the thioether 22 that was formed during the reaction. Ultimately, the efficient synthesis of salacinol (13, 75% overall yield) was achieved by reacting the *p*-methoxybenzyl (PMB)-protected thioether 24, instead of the benzyl-protected thioether 22, with the cyclic sulfate 20 and the problematic hydrogenolysis step was eliminated as, in this case, all the protecting groups were readily removed using aqueous trifluoroacetic acid (TFA), as shown in Scheme 4b.

$$P_{10}$$
 P_{10}
 P_{10}
 P_{20}
 P_{10}
 P_{10}

 P_1 and P_2 = protecting groups

Figure 6. General strategy for the alkylation at the heteroatom of a hetero anhydroalditol.

2.3. Heteroatom analogues of salacinol

The subsequent syntheses of salacinol and its analogues have incorporated many improvements especially with respect to the syntheses of the respective cyclic sulfates derived from a variety of polyhydroxylated heterocyclic ethers that have been employed as coupling partners. However, the key strategy for the formation of the zwitterionic alkylated product (Fig. 6), that is, the nucleophilic ring opening of a protected cyclic sulfate by the heteroatom of a protected or unprotected hetero anhydroalditol, developed by Yuasa et al.³⁰ and our group, ^{31,34} remains unchanged.

2.3.1. Nitrogen analogues. In 2001, we reported the syntheses of two nitrogen analogues (25, ghavamiol and 26) of salacinol. Compounds 25 and 26 were synthesized by reactions of the iminoarabinitol 27^{36} with the cyclic sulfate 20 and the iminoarabinitol 28^{37} with the cyclic sulfate 21, respectively, as shown in Scheme 5. Compounds 25 and 26 were tested against three glucosidase enzymes, namely glucoamylase G2, porcine pancreatic α -amylase (PPA), and barley α -amylase

Scheme 5.

(AMY1), and the effects were compared to those of salacinol (13). It was observed that the ammonium compounds 25 and 26 were not active against AMY1 and PPA at concentrations of 5 mM compared to salacinol (13) that showed stronger inhibition against AMY1 and PPA ($K_i = 15 \pm 1$ and $10 \pm 2 \,\mu\text{M}$, respectively). Salacinol (13) showed weak inhibition of glucoamylase G2 ($K_i = 1.7 \,\text{mM}$) whereas the ammonium compounds 25 and 26 inhibited glucoamylase with K_i values in the range ~ 10 -fold higher than salacinol.

Screening of salacinol and the nitrogen analogues against human pancreatic α -amylase (HPA) showed that compound **26** was an inhibitor of this enzyme with a K_i value of 0.4 mM compared to salacinol that showed stronger inhibition with a K_i value of 75 μ M. In contrast, ghavamiol (**25**) was not active against this enzyme.³⁸

Ghavamiol (25) was also independently synthesized by Muraoka et al.³⁹ and was tested for its α-glucosidase inhibitory activities against rat small-intestinal disaccharidases; maltase, sucrase, and isomaltase. When compared to salacinol (13), the inhibitory activities of ghavamiol (25) against these enzymes decreased considerably. In approaching the target 25, the same ring-opening strategy using the iminoarabinitol 27 and the cyclic sulfate 18 was followed; however, unlike the previous syntheses of salacinol^{30,31,34} and ghavamiol³⁵ in which the respective cyclic sulfates 18 and 20 were obtained from L-glucose, an optimized synthesis of the cyclic sulfate 18 starting from a benzylidene-protected diol that was readily obtained from D-glucose^{40–42} was presented (Scheme 6).

It is of interest that, at a concentration of 1 mmol/L, ghavamiol (25) inhibited \sim 96% of the recombinant lysosomal acid α -glucosidase (GAA) activity, whereas this enzyme was relatively insensitive to the α -glucosidase inhibitor, acarbose.⁴³ In addition, ghavamiol showed

selective inhibition of the activity of GAA in the detergent extract of human polymorphonuclear leukocytes (PMNs), which also contains a second acid α -glucosidase known as maltase glucoamylase (RAAG). Inhibition studies in which recombinant GAA was added to the PMN extract showed that all of the recombinant enzyme was inhibited by 1 mmol/L ghavamiol in the presence of all the other components from the PMN lysate. ⁴³ This selective inhibition of the enzyme GAA has been used to advantage in the design of a diagnostic test for Pompe's disease.

In all of the above syntheses of salacinol and related compounds, the alkylation reactions between the various 1,4-anhydroarabinitol moieties and the cyclic sulfates proceeded stereoselectively irrespective of the reaction conditions. The trans relationship between C-5 and C-1' was assigned by means of NOE correlations observed between H-4 and H-1' in NOESY experiments.

2.3.2. Selenium analogues. We have also described the convenient syntheses of two selenium analogues of salacinol, selenosalacinol (*trans-29*), and its stereoisomer $30.^{44-46}$ Compounds *trans-29* and 30 were tested against different glycosidases and the results were compared to those of salacinol (13). Selenosalacinol (*trans-29*, named blintol) was found to be a weak inhibitor of glucoamylase G2 ($K_i = 0.72$ mM) and showed no significant inhibition of AMY1, HPA, 38 or PPA. The stereoisomer 30 of blintol showed no significant inhibition of glucoamylase G2, HPA, 38 PPA, or AMY1. The activity of blintol (*trans-29*), ghavamiol (25), and salacinol (13) against recombinant human intestinal maltase glucoamylase (MGA) will be described later.

The original synthesis of blintol began with the preparation of the per-benzylated selenoether **31** that was derived from the intermediate **32** by its conversion into the corresponding dimesylate **33**³⁷ and subsequent reaction with freshly prepared Na₂Se (Scheme 7a). The

Scheme 7a.

intermediate 32 was prepared from L-xylose in four steps using a literature method.³³ The coupling reaction was again based on similar strategies developed for the synthesis of salacinol and related compounds, except that the selenoether 31 and the cyclic sulfate 34³⁰ were used as coupling partners.

Unlike the reported reactions of an anhydrothioarabinitol or iminoarabinitol with various cyclic sulfates (e.g., Schemes 2–6), the reaction of the selenoether 31 with the cyclic sulfate 34 yielded the alkylated selenonium ion 35 as a 3:1 mixture of diastereomers at the stereogenic selenium center (Scheme 7a). The major isomer was identified, by means of 1D-NOESY experiments, as the diastereomer with a trans relationship between C-5 and C-1'. Hydrogenolysis of the mixture (35) gave a mixture of diastereomers (trans-29 and cis-29) that,

upon precipitation from MeOH, yielded the pure blintol (*trans*-29). The stereoisomer 30 was prepared in a similar fashion from the selenoether 31 and the cyclic sulfate 21 (Scheme 7b).

We have also achieved an efficient synthesis of blintol using a PMB-protected selenoether (36) and the benzylidene-protected cyclic sulfate (20) (Scheme 8).⁴⁷ This synthesis required, in turn, an optimized synthesis of the selenoether 36, which featured the use of boric acid in the acid-catalyzed acetylation of L-xylose to improve the furanoside:pyranoside ratio⁴⁸ and the use of an *n*-pentenyl glycoside in an intermediate step⁴⁹ (Scheme 9). In addition, the benzylidene-protected cyclic sulfate 20, the other coupling partner, was successfully prepared from D-glucose instead of expensive L-glucose, as depicted in Scheme 10.

Scheme 7b.

Scheme 8.

Scheme 9.

D-glucose
$$\frac{\text{ref.31}}{4 \text{ steps}}$$
 Ph O OBn $\frac{\text{ref.44}}{3 \text{ steps}}$ O OBn $\frac{\text{H}_2, \text{Pd/C}}{OBn}$ OBn $\frac{\text{H}_2, \text{Pd/C}}{OBn}$

Scheme 10.

2.4. Five-membered ring analogues with different sugar stereochemistries

To probe the importance of the configurations of the stereogenic centers in the heterocyclic ring of salacinol (13), we have described the synthesis of an anhydrothio-D-xylitol analogue (37) of salacinol and also the corresponding nitrogen congener (38) (Fig. 7).⁵⁰ In these compounds, the stereochemistries at C-2 and C-3 were both inverted compared to salacinol. As shown in Scheme 11, the synthesis started with the preparation of the respective coupling partners, anhydrothio- (39) and imino- (40) xylitols, from a common precursor 41 that was readily prepared in five steps from L-arabinose by use of a literature procedure.⁵¹

Enzyme inhibition studies indicated that the sulfonium analogue 37 inhibited, albeit weakly when com-

Figure 7. An anhydrothio-D-xylitol analogue (37) of salacinol and the corresponding nitrogen congener (38).

pared to salacinol (13), AMY 1 and PPA with K_i values of 109 ± 11 and $55 \pm 5 \,\mu\text{M}$, respectively. However, the ammonium-ion analogue 38 showed no significant inhibition against these enzymes.

The syntheses of anhydrothio-D-lyxitol (42) and anhydrothio-D-ribitol (43) analogues of salacinol (Fig. 8), in which the stereochemistries at C-3 and C-2, respectively, are inverted compared to salacinol, have also been reported. These compounds were synthesized by use of PMB-protected thioethers 44 and 45, respectively, and the cyclic sulfate 20. The PMB-protected thioether 44 was conveniently prepared from the intermediate 46, as shown in Scheme 12. A large-scale synthetic route for the key intermediate 46 was developed starting from cheaper D-lyxose, as opposed to the literature method starting from L-lyxose, by employing a Mitsunobu reaction as a key step to yield the required stereochemistry at C-4 (Scheme 12).

The thioether **45** was obtained from D-ribose by a literature route similar to the above protocol described for the thioether **44**. ⁵⁴ Enzyme inhibition studies of **42** and **43** against MGA indicated that they were not effective inhibitors against MGA, whereas salacinol inhibited this enzyme with a K_i value of 0.19 μ M. ⁵⁵ Hence, it was concluded that the D-arabinitol configuration in the heterocyclic ring displayed by salacinol is critical for its activity.

Scheme 11.

Figure 8. Anhydrothio-p-lyxitol (42) and anhydrothio-p-ribitol (43) analogues of salacinol and PMB-protected precursors to these structures, 44 and 45.

With the aim of deriving potential inhibitors of UDP-galactopyranose mutase, we have also synthesized the related sulfonium salts (47 and 48) with 1,4-anhydro-p-galactitol moieties,⁵⁶ as well as the corresponding nitrogen (49 and 50) and selenium (51 and 52) analogues (Fig. 9).⁵⁷ The syntheses of the nitrogen and selenium

analogues (49–52) made use of the respective coupling partners 53 and 54. The synthetic highlight of this report was the synthesis of 54 from D-glucose by a multi-step synthetic route that involved selective deprotection of the anomeric pentenyl glycoside in the presence of acid labile PMB ethers using NBS (Scheme 13); the corresponding imino-D-galactitol (53) was prepared by a slightly modified literature method. Enzyme inhibition studies of the final zwitterionic compounds (47–52) indicated that they did not inhibit UDP-galactopyranose mutase.

2.5. Six-membered ring analogues

To study the effect of the size of the heterocyclic ring on glycosidase inhibitory activity, we have synthesized a series of six-membered ring analogues (55–59) of salacinol that were modeled after miglitol. ⁵⁹ Each of these compounds has a six-membered, cyclic alditol structure, with a positively charged heteroatom (sulfur or selenium or nitrogen) and also contains either a D- or L-erythritol-3-sulfate side chain (Fig. 10). The coupling reactions

Figure 9. 1,4-Anhydro-D-galactitol analogues synthesized as potential inhibitors of UDP-galactopyranose mutase.

Scheme 13.

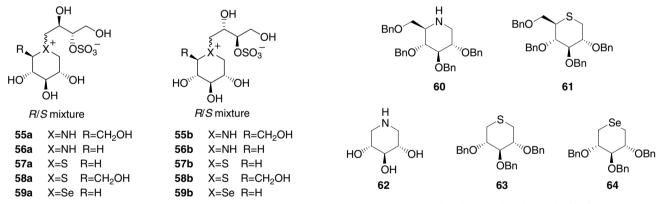


Figure 10. Six-membered ring analogues of salacinol.

were performed by the standard protocol using the respective heterocyclic ethers 60–64 (Fig. 11) and the cyclic sulfates 20 and 21. Unlike the five-membered ring analogues, the final products were obtained as mixtures of epimers, separable in some cases, differing only at the configurationally stable sulfur or selenium atoms. The required starting materials were prepared using literature methods (62)⁶⁰ or readily obtained in one (60⁶¹ and 61²¹) or two steps (63 and 64)⁶² from known intermediates. An important conclusion derived from the enzyme inhibition studies was that the five-membered ring of salacinol (or heteroanalogues) is a necessary and key structural determinant for its activity, as all the six-membered ring

analogues were inactive against MGA or HPA.

Figure 11. Heterocyclic ethers used in the synthesis of 55-59.

Recently, Gallienne et al.⁶³ reported the syntheses of two series of five- and six-membered ring analogues of salacinol involving thioanhydro alditol moieties with *erythro*, D, L-threo, xylo, ribo, D-arabino, and D-manno configurations (65–74, Fig. 12), including the previously reported six-membered ring analogue 57b.⁵⁹ Apart from synthesizing these zwitterionic compounds, by following the general strategy for the coupling reaction, they have reported an alternative synthesis of the L- and D-cyclic sulfates (18 and 21) from a common intermediate (75) that was readily obtained from 4,6-O-benzylidene-D-glucose (76).⁴⁰ This method exploits two one-pot procedures to obtain the required cyclic sulfates from 4,6-O-benzylidene-D-glucose (Scheme 14). The synthesis began

Figure 12. Analogues of salacinol synthesized by Gallienne et al. 63

Scheme 14.

with the periodate oxidation of benzylidene-protected D-glucose 76, followed by the reduction of the resulting aldehyde with NaBH₄ in one pot. The benzylidene-protected diol 75 was transformed into the required D-cyclic sulfate 21 using a one-pot procedure. In the case of the L-cyclic sulfate (18), the diol (75) was first protected as an isopropylidene ketal and the benzylidene-protecting group was then selectively removed. The resulting iso-

propylidene-protected diol was subjected to the one-pot procedure, as before, to give the L-cyclic sulfate (18).

These compounds (65–74 and 57b) were screened against six different commercial glycosidases including the rice-derived α -glucosidase. All compounds from the five-membered ring series (65–70) that are missing the hydroxymethyl group at C-4 when compared to salacinol were inactive against the α -glucosidase from rice,

whereas, salacinol (13) had been shown previously to exhibit a strong inhibitory effect toward this enzyme (IC₅₀ = 1.1×10^{-3} mM). ⁶⁴ Hence, it was concluded that the hydroxymethyl group of salacinol is essential for its inhibitory activity toward this particular enzyme. The six- and seven-membered ring analogues exhibited only very weak activities or no activity against the glycosidases tested, except for the xylitol analogue **57b** (R/S mixture), which showed an inhibitory effect against β -glucosidase from almond. Based on this study and our enzyme inhibition data for the various six-membered ring analogues (**55–59**), ⁵⁹ we conclude that, in general, the ring expansion of the heterocycle displayed in salacinol leads to loss of inhibitory activity.

This conclusion was corroborated, when Gallienne et al. 65 synthesized a new series of salacinol analogues (77–80, Fig. 13) containing six-membered ring nitrogen heterocycles and tested them against six commercial glycosidases including rice α -glucosidase and baker's yeast α -glucosidase. As observed with the other six-membered ring analogues, none of the synthesized ammonium salts were active against the glycosidases tested.

The synthetic route involved the facile synthesis of two, enantiomerically pure, iminosugars (81 and 82) and their further transformation into the ammoniumion analogues (77–80) by reacting them with cyclic sulfates 18 and 21 (Scheme 15). The syntheses began with the preparation of the novel isoxazoline cyclic sulfates, 83 and 84, in three steps starting from the optically pure alkene 85⁶⁶ and the nitro compound 86.⁶⁷ Hydrogenolysis of the isoxazolines 83 and 84, followed by acid treatment of the resulting zwitterionic intermediates, gave the iminosugars 81 and 82, respectively. Alkylation reactions of these iminosugars with the cyclic sulfates 18 and 21, followed by the removal of the protecting group, gave the final zwitterionic ammonium salts.

Recently, the synthesis of related six-membered ring sulfonium ions (87–90), containing 1,5-anhydro-5-thio-L-fucitol as a core structure, as potential α -L-fucosidase inhibitors was described. These compounds were obtained from the alkylation reactions of 1,5-anhydro-5-thio-L-fucitol (91) with the cyclic sulfates, 20 and 21, or various alkyl halides (Scheme 16).

The starting material, 1,5-anhydro-5-thio-L-fucitol (91), was readily prepared in few steps from a selectively

Scheme 15.

protected mannopyranose derivative (92),^{70,71} as shown in Scheme 17. The synthesis began with the nucleophilic addition of a methyl group to the intermediate 92 via a Grignard reagent, and then, sodium periodate oxidation and sodium borohydride reduction reactions were performed sequentially to get the diol 93. This diol was further transformed into the required anhydrothiofucitol derivative (91) by forming the corresponding dimesylate followed by its reaction with sodium sulfide.

Figure 13. Salacinol analogues synthesized by Gallienne et al. 65 containing a six-membered ring nitrogen heterocycle.

Scheme 16.

Scheme 17.

2.6. Chain-modified analogues

Based on the fact that kotalanol (14), with a longer polyhydroxylated side chain, has been reported to possess greater inhibitory activity against certain glucosidase enzymes when compared to salacinol (13), 23 much attention has been directed toward the synthesis of acyclic chain-modified analogues of salacinol. Accordingly, several chain-modified analogues have been synthesized and tested for their glucosidase inhibitory properties. These modifications include the extension of the acyclic chain, the change in the position or complete removal of the sulfate group, and the introduction of acyclic chains that are missing one or two hydroxyl groups when compared to salacinol.

2.6.1. Chain-extended analogues. In 2006, we reported the synthesis and enzyme inhibitory activity of four chain-extended homologues (94–97) of salacinol containing polyhydroxylated, acyclic chains of 5- and 6-carbons, differing in stereochemistry at the stereogenic centers. ^{72,55} The syntheses involved the reactions of anhydro-4-thio-p-arabinitol (22) with cyclic sulfate derivatives of different monosaccharides (98–101, ^{73–76} Fig. 14). Deprotection of the benzyl groups followed by reduction of the hemiacetal intermediates (102–105) gave the final compounds 94–97 (Scheme 18). With the exception of compound 94, all the other compounds 95–97 inhibited MGA, with K_i values of 0.25, 0.26, and 0.17 μ M, respectively. By comparison of the stereo-

Figure 14. Cyclic sulfates used in the synthesis of chain-extended homologues of salacinol containing polyhydroxylated, acyclic chains of 5- and 6-carbons.

Scheme 18.

chemistry of the stereogenic centers in the extended acyclic chain and the observed inhibitory properties of the four compounds, it was concluded that the *R*-stereochemistry of the C-4' stereogenic center is critical for activity. The hemiacetal derivatives 102–105 were also tested for their glycosidase inhibitory properties; however, these compounds were not active against MGA.

Recently, we also reported a series of six chainextended analogues (106–111, Fig. 15) with heteroatom variations, containing polyhydroxylated, extended acyclic chains of 6-carbons.⁷⁷ These analogues were prepared by coupling the PMB-protected D- and Lanhydroseleno-, anhydrothio-, and iminoarabinitols (24, 36, and 112–115, 34,47,46 Fig. 15) with the cyclic sulfate **116** that was readily prepared in three steps from D-sorbitol, as shown in Scheme 19.

The enzyme inhibition studies against MGA revealed that the sulfur analogue 107 with the D-arabinitol configuration in the heterocyclic ring did not inhibit MGA, while the corresponding selenium (106) and nitrogen (108) analogues inhibited MGA with K_i values of 41 and 26 μ M, respectively. Interestingly, the sulfur and nitrogen congeners with the L-arabinitol configuration in the heterocyclic ring, 110 and 111, were also active, with K_i values of 25 and 5 μ M, respectively.

Recently, the syntheses of the acyclic, chain-modified, sulfur (117) and selenium (118) analogues (Fig. 16), with an extended polyhydroxylated acyclic chain of

Figure 15. Chain-extended analogues of salacinol with heteroatom variations, containing polyhydroxylated, extended acyclic chains of 6-carbons (106–111), and precursors used in their synthesis (24, 36, 112–115).

Scheme 19.

Figure 16. Sulfur (117) and selenium (118) analogues of salacinol, with an extended polyhydroxylated acyclic chain of 6-carbons and the cyclic sulfate (119) used in their synthesis.

6-carbons, have been reported.⁷⁸ These analogues were synthesized using the cyclic sulfate **119**. The synthesis of **119** was achieved by the selective acetolysis of a suitably protected D-mannitol derivative followed by 1,3-cyclic sulfate formation, as shown in Scheme 20. The main reason for the use of the cyclic sulfate **119** was to maintain the stereochemistry at C-2' and C-3' as in salacinol (**13**), and the stereochemistry at C-4' as in the other active chain-extended analogues (**95–97**) that were reported previously.⁷² Enzyme inhibition studies indicated that **117** and **118** were effective inhibitors of MGA, with K_i values of 0.65 and 0.14 μ M, respectively.

We have developed an alternative synthetic route to the active chain-extended analogues 95 and 96 (Scheme 21) that involves the use of 1,3-cyclic sulfates (120 and 121) protected with an acid sensitive, butane diacetal (BDA)-protecting group compared to the previous synthetic route, which used 1,3-cyclic sulfates with benzyl-protecting groups (Scheme 18 and Fig. 14). This new strategy eliminates the difficulties of the previous synthetic route with respect to the removal of the benzyl-protecting groups, which limited our access to the corresponding selenium analogues; hence, by use of BDA-protected cyclic sulfates 120 and 121, the corresponding

selenium analogues, **122** and **123**, were synthesized successfully (Scheme 21). ⁷⁹ In addition, the corresponding nitrogen analogues, **124** and **125**, were also synthesized. The cyclic sulfates **120** and **121** were readily obtained in two steps from benzyl β -D-galactopyranoside (**126**)⁸⁰ and benzyl β -D-glucopyranoside (**127**), ⁸¹ respectively, as shown in Scheme 22.

Enzyme inhibition studies indicated that the selenium analogues 122 and 123 inhibited MGA, both with K_i values of 0.10 μ M. These compounds constitute the most active to date in this series of zwitterionic glycosidase inhibitors. However, the nitrogen analogues 124 and 125 were less active compared to the corresponding sulfur and selenium analogues, with K_i values of 35 and 8 μ M, respectively. These data corroborate our previous conclusion with other derivatives (95–97 and 117–118), T2.78 that is, the configuration at C-3′ does not appear to be critical for inhibitory activity, as each of the pairs with enantiomeric configurations at C-3′, 122 and 123; 124 and 125 has similar K_i values.

Comparison of the K_i values of the sulfonium and selenonium chain-extended analogues (94–97, 106, 107, 117, 118, 122, and 123) against MGA indicated that for both the 5- and 6-carbon chain-extended series, the S-configuration at C-2' and the R-configuration at C-4' are critical for activity (the highlighted portions in Table 2). T2,77–79,55 Furthermore, the configuration at C-3', bearing the sulfate moiety, was found to be unimportant. The configurational requirement at C-5' was not clear because the trend was found to depend on the heteroatom. However, the overall conclusion was that the effect of the acyclic chain extension did not confer any dramatic change on inhibitory properties for this particular enzyme, because the K_i values of the active chain-extended analogues were in a similar range to those of salacinol (13) and blintol (trans-29, 0.49 μ M).

Scheme 21.

Scheme 22.

2.6.2. Deoxy-salacinol analogues. Recently, Muraoka et al. 82 reported the syntheses and glycosidase inhibitory properties of three deoxy-salacinols lacking one (128 and 129) or two polar (130) substituents in the side chain of salacinol (13); the syntheses made use of the thioara-

binitol **16** and the cyclic sulfates **131–133** as coupling partners (Fig. 17). In addition, an alternative synthetic route to **16** was achieved via the selective *tert*-butyldimethylsilyl (TBDMS)-protection of the primary hydroxyl group of the intermediate **134** that was prepared from

Table 2. K_i values against MGA for various chain-extended analogues of salacinol with an identical anhydro-D-arabinitol configuration in the heteroalditol ring and acyclic chains of 5- or 6-carbons with different stereochemistry at the stereogenic centers

	Stereoch					
Inhibitor	- Clereocii	emistry at the in the acyc	<i>K</i> _i (μΜ)	Reference		
	C-2'	C-3'	C-4'	C-5'	Α (μινι)	Helefelice
94 ^a	S R S		NA	NI)	
95 ^a	S	R	R	S	0.25	70
96 ^a	s	S	R	S	0.26	> 72
97 ^a	S	S	R	NA	0.17	J
106 ^b	R	S	R	R	41	l 77
107 ^a	R	s	R	R	NI	<i>''</i>
117 ^a	S	S	R	R	0.65	78
118 ^b	s	S	R	R	0.14	, ,
122 ^b	s	R	R	s	0.10	79
123 ^b	S	S	R	S	0.10] /9

NA = not applicable; NI = no inhibition.

D-xylose in two steps (Scheme 23). Benzylation of the C-3 hydroxyl of **135**, followed by removal of the other protecting groups and reaction with mesyl chloride, gave the dimesylate **136**. Compound **136** was then converted into the thioarabinitol **16** in three steps, as shown in Scheme 23. The cyclic sulfate **133** was synthesized from 1,3-*O*-benzylidene-L-erythritol (**75**) via deoxygenation of

the xanthate intermediate **137**, as shown in Scheme 24, while the other cyclic sulfates (**131** and **132**) were synthesized using modified literature methods. 83–88

Enzyme inhibition studies indicated that compounds 129 and 130 having acyclic chains that lack the hydroxyl group at C-2′, compared to the acyclic chain of salacinol (13), were not effective inhibitors of intestinal α -glucosidases in vitro. However, compound 128, lacking a hydroxymethyl group at C-3′, retained some inhibitory activity against sucrase (IC₅₀ = 780 μ M). These results suggested that both the hydroxymethyl group at C-3′ and the hydroxyl group at C-2′ with an S-configuration are essential for α -glucosidase inhibitory activity.

2.6.3. Frame-shifted analogues. To probe the effect of the positioning of the sulfate group in the acyclic chain, we have synthesized the chain-modified analogues (138– 141, Fig. 18) with an extended chain of 5-carbons in which the position of the sulfate moiety was shifted from C-3' to C-4' compared to salacinol (13) and other chainextended analogues (Table 2).89 The target compounds were synthesized by the reactions of the heteroalditols 24 and 36 with the cyclic sulfates 142 and 143, followed by removal of the protecting groups. The synthetic highlight was that three of the four required coupling partners, namely, the seven-membered cyclic sulfate (142), the PMB-protected thioether (24), and selenoether (36), were derived from a common diol precursor 144, as shown in Scheme 25. The other cyclic sulfate 143 was synthesized in a similar manner starting from the

Figure 17. Synthetic deoxy-salacinols lacking one (128 and 129) or two polar (130) substituents in the side chain and the cyclic sulfates (131–133) employed as coupling partners.

^aSulfur analogues.

^bSelenium analogues.

Scheme 24.

Figure 18. Chain-modified analogues in which the position of the sulfate moiety is shifted from C-3' to C-4'.

enantiomeric diol **145** (Scheme 25). The diols **144** and **145** were synthesized in turn from D- or L-xylose according to the procedure developed recently in our laboratory. ⁴⁷ The target compounds **138** and **139** showed inhibition of MGA with K_i values of 20 and 53 μ M,

respectively. In contrast, compounds **140** and **141** were not active against MGA.⁸⁹

2.6.4. De-O-sulfonated analogues. Very recently, Tanabe et al. ⁹⁰ reported the de-O-sulfonated analogues of salacinol (**146** and **147**, Fig. 19) and studied the role of

Figure 19. De-O-sulfonated analogues of salacinol (146 and 147) and their inhibitory activities.

the sulfate anion moiety in the acyclic chain of salacinol (13) on inhibitory activity. The de-O-sulfonated salacinol 146 was synthesized by de-O-sulfonation of salacinol (13) using methanolic hydrogen chloride, and compound 147 was obtained by simply exchanging the monomethyl sulfate counterion of 146 with chloride counterion using an ion exchange resin. The enzyme inhibition studies indicated that the internal sulfate counterion is not essential for inhibitory activity as the de-O-sulfonated analogues (146 and 147) showed almost equal inhibitory activities against intestinal α-glucosidases (in vitro) when compared to salacinol (13), irrespective of the external counterion (Fig. Presumably, Nature biosynthesizes the sulfated analogues for purposes of stability, solubility, or more facile transport into cells.

2.7. Carboxylate analogues

In an attempt to understand the role of the sulfate group present in the acyclic chain of ghavamiol (25), we have synthesized an unusual class of amino acids 148 and 149 (Fig. 20). 91 These analogues were considered to be

Figure 20. Carboxylate analogues of ghavamiol (148 and 149) and compounds used in their preparation (150, 151, and 154).

the carboxylate analogues of ghavamiol. Compound 148 was found to be an inhibitor of MGA with a K_i value of 21 μ M. In addition, this compound was also active against *Drosophila melanogaster* Golgi mannosidase II (dGMII), with an IC₅₀ of 0.3 mM. Compound 149 was not active against either MGA or dGMII. The crystal structure of 148 bound in the active site of dGMII indicated that the hydroxyl groups from the acyclic chain and also the carboxylic acid group form extensive contacts with both side chains and water molecules in the active site.⁹¹ Previously, the role of the sulfate group on glycosidase inhibitory activity was inferred by Yuasa et al.;⁹² the docking of salacinol into the binding site of glucoamylase indicated close contacts between the sulfate ion and Arg305.

The key step in the synthesis of compound 148 was the reaction of the iminoarabinitol 150 with the epoxide 151 that was prepared from vitamin C using a simplified literature procedure. Debenzylation of the coupled product 152 followed by stereoselective catalytic reduction of the double bond of the L-ascorbic acid moiety produced the lactone 153. The desired amino acid 148 was derived from the lactone 153 by hydrolysis followed by ion exchange resin treatment, as depicted in Scheme 26. Compound 149 was prepared in a similar fashion using the iminoarabinitol 154.

Very recently, using the epoxide 155⁹⁴ and the thioethers 22 and 23, the sulfonium-ion analogues, 156 and 157 (Fig. 21), with an internal carboxylate counterion were also synthesized by our group. 95 Compound

Figure 21. Carboxylate analogues of salacinol (156 and 157) and the protected epoxy ester 155 used in their synthesis.

156, with the D-arabinitol configuration in the heterocyclic ring, inhibited MGA with a K_i value of 10 μ M, whereas compound **157** did not show significant inhibition against MGA.

2.8. Phosphate derivatives

We have synthesized the phosphate derivatives related to salacinol (158 and 159, Fig. 22) that are missing the hydroxyl groups in the side chain and screened them against MGA. ^{26,96} Compounds 158 and 159 were considered to be the prototypes of the phosphate analogue 160 that has the exact structural features of salacinol except for the phosphate group that replaces the sulfate moiety at C-3' (Fig. 22). However, compounds 158 and 159 were not effective against MGA. We have also attempted the synthesis of the exact phosphate analogue of salacinol 160 using the cyclic phosphate 161 or 162 and the thioether 22. ^{26,96} Unfortunately, the coupling reaction did not proceed as planned even after several attempts under various reaction conditions.

2.9. 2-Amido- and 2-amino derivatives

The syntheses of 2-acetamido (163) and 2-amino (164) derivatives of salacinol were also attempted (Fig. 23). However, it was found that these derivatives were unstable and underwent ring-opening reactions to give the acyclic amido sulfate 165 and ammonium sulfate 166.

The synthesis started with the key azido intermediate 167, obtained from 1,4-anhydro-4-thio-p-ribose. ⁵⁴ As depicted in Scheme 27, sequential reduction and acetylation of the intermediate 167, followed by a change of protecting groups, gave the required coupling partner 168 for the synthesis of the acetamido derivative 163. However, the coupling reaction between 168 and the cyclic sulfate 20 did not lead to the desired product, giving instead the ring-opened product, an acyclic amido sulfate 165.

Figure 23. 2-Acetamido (163) and 2-amino (164) analogues of salacinol.

This ring-opening reaction can be attributed to the nucleophilic attack of the amide oxygen on C-1 of the ring (Scheme 27). In an attempt to avoid this intramolecular reaction, the azido intermediate 169, derived from the intermediate 167, was coupled directly with the cyclic sulfate 20 and indeed, the alkylated product 170 was formed in this case. However, during reduction of the azido group by hydrogenation, the ring-opened product 166 was formed, presumably through nucleophilic participation of the free amine (Scheme 27).

It is of interest that the corresponding nitrogen analogues 171–176 are stable (Fig. 24). 98

2.10. Other salacinol-related analogues

Salacinol analogues (177–184, Fig. 25) based on novel anhydroseleno- and anhydrothio-allitols (185–188), derived from D-gulono-γ-lactone (189) and L-ascorbic acid (190), have also been synthesized. Protection of the hydroxyl groups in D-gulono-γ-lactone, loo followed by sequential reduction and mesylation reactions, loo produced the dimesylate that, upon treatment with selenium metal and sodium borohydride or Na₂S, produced the anhydroseleno- and anhydrothio-D-allitols, 185 and 186, respectively, as shown in Scheme 28.

The enantiomeric allitols (187 and 188) were also synthesized in a similar manner (Scheme 29), except

Figure 22. Phosphate derivatives related to salacinol (158 and 159) and the cyclic phosphate derivatives (161 and 162) used in the attempted synthesis of the exact phosphate analogue of salacinol, 160.

Scheme 27.

Figure 24. Analogues of ghavamiol with a nitrogen substituent at C-2.

that the syntheses started from L-ascorbic acid (190) instead of expensive L-gulono- γ -lactone. These allitols were subjected to coupling reactions with the cyclic sulfates 20 and 116, and the coupled products were

then deprotected to obtain the zwitterionic compounds (177–184). Enzyme inhibition studies of these compounds indicated that they were not active against MGA.

Figure 25. Salacinol analogues (177–184) derived from D-gulono-γ-lactone and L-ascorbic acid.

Scheme 28.

Scheme 29.

Using the cyclic sulfate 116, the synthesis of another series of chain-extended analogues (191–194, Fig. 26), containing a heterocyclic five- and six-membered ring

core with polyhydroxylated acyclic chains of six carbons, was reported by our group. 103 The salient feature of this report was that all of the required

Figure 26. Chain-extended analogues (191–194) containing a heterocyclic five- and six-membered ring core with polyhydroxylated acyclic chains of six carbons, which were synthesized from 195 to 198.

Figure 27. 5-Thioglycopyranosides 199 and 200 and the corresponding sulfonium-ion derivatives 201-204.

Figure 28. Sulfonium (205 and 206) and ammonium ion (207 and 208) analogues of salacinol, containing an additional hydroxymethyl group at C-1, which were obtained from thioether 209 and iminoalditol 210.

coupling partners (195–198) were synthesized from D-mannose.

Tian et al.¹⁰⁴ have reported the efficient synthesis of 5-thioglycopyranosides, **199** and **200**, starting from D-xylose and D-ribose, respectively. These compounds were further transformed into the sulfonium derivatives **201–204** using the cyclic sulfate **21** and methyl iodide (Fig. 27). In this report, they also proposed a simple ¹³C NMR-based method to determine the stereochemistry at the stereogenic sulfonium center of the six-membered ring sulfonium salts.

Recently, we reported the synthesis of the sulfonium (205 and 206) and ammonium ion (207 and 208) analogues of salacinol, containing an additional hydroxymethyl group at C-1, using the thioether (209) and iminoalditol (210) (Fig. 28). ¹⁰⁵

Our recent additions to the repertoire of salacinol-related compounds include a series of S-alkylated sulfonium ions with varying alkyl chains (211–219, Fig. 29). These lipophilic compounds were shown to be effective inhibitors of MGA, with K_i values ranging from 6 to 75 μ M.

3. Crystallographic and NMR/molecular modeling studies of salacinol and its analogues

We have investigated the crystal structures of salacinol (13), blintol (*trans-29*), ghavamiol (25), and also the stereoisomers of salacinol and blintol, 17 and 30, respectively, bound in the active site of Golgi α -mannosidase II. 107 Although these compounds show poor

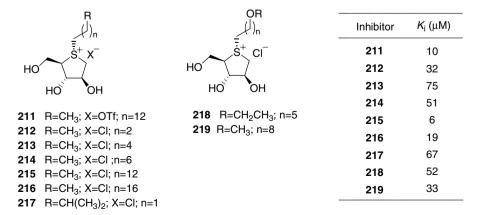


Figure 29. S-Alkylated sulfonium ions with varying alkyl chains (211-219) and their inhibitory activities against MGA.

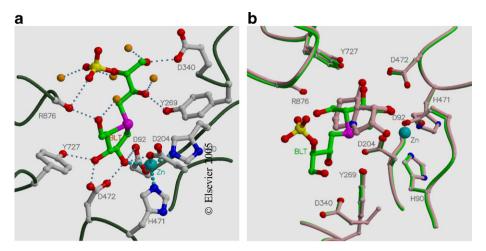


Figure 30. (a) View of the GM II enzyme interactions with compound 30, (b) overlay of 30 and swainsonine in the active site of GM II. Reprinted with permission from Ref. 107.

inhibitory activity against GM II (in the millimolar range), they form critical interactions with the active-site carboxylate Asp 204 (Fig. 30a), as also observed in the crystal structure of swainsonine (2) bound in the active site of GM II. For example, the selenonium center in compound 30 is located in a similar position to the nitrogen atom in swainsonine (2). The hydroxyl groups OH-2 and OH-3 overlap very favorably with the OH-1 and OH-8 hydroxyl groups of swainsonine (Fig. 30b).

The transient positive charge of the transition state in the glucosidase-mediated hydrolysis reaction is presumably mimicked by the permanent positive charge provided by the sulfonium, ammonium, or selenonium ions in these compounds. The interactions between these positive centers with the side chain of Asp 204 are indicative of the electrostatic stabilization provided by the enzyme. All these compounds form T_5 coordination with the active-site Zn atom whereas swainsonine forms T_6 coordination with zinc. Hence, the lower inhibitory activities observed for these compounds might be a consequence of the lack of T_6 coordination with the active-site Zn atom.

A combined saturation transfer difference NMR (STD-NMR)/molecular modeling protocol applied to the dGMII-salacinol 13 complex was also published by our group. The experimental STD-NMR results together with theoretical STD values, calculated with the CORCEMA-ST program (complete relaxation and conformational exchange matrix analysis of saturation transfer), 109,110 correlated well with the lowest-energy binding modes shown by molecular modeling. Of the different conformers obtained using Auto Dock 3.0, conformer E₄ exhibited the most favorable docked energy (-11.71 kcal mol⁻¹). This conformation was similar to that of salacinol (13) in the complex with dGMII in the crystal structure (Fig. 31).

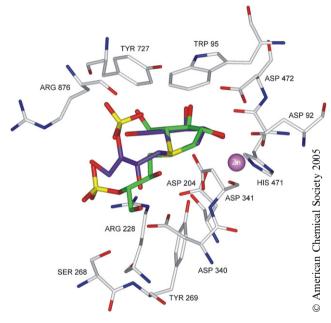


Figure 31. The predicted structure (green carbon atoms) obtained by Auto Dock 3.0, superimposed on the crystal structure of salacinol (13) (amethyst carbon atoms) in the active site of dGMII. Reprinted with permission from Ref. 108

4. Concluding remarks

It is of interest that studies with herbal extracts of *S. reticulata* with rats have shown control of blood glucose levels after a carbohydrate meal. Studies have also shown that the oral ingestion of an extract from a *S. reticulata* trunk at a dose of 5000 mg kg⁻¹ had no serious acute toxicity or mutagenicity in rats. It is also noteworthy that a double-blind study of the effects of the extract from *S. reticulata* on human patients with type II diabetes mellitus has shown that the extract is an effective treatment of type II diabetes with side effects

Table 3. Glycosidase inhibitory activities of salacinol and related compounds

Inhibitor		β-Glcase ^a	α-Galase ^b	β-Galase ^c	α-Manase ^d	Ref.							
	Rat intestinal disaccharidase K_i (μ M)	Gluco-amylase G2 K_i (mM)	PPA K_i (mM)	AMY1 K _i (mM)	MGA K _i (μM)	HPA K _i (mM)	Rice IC ₅₀ (mM)	Baker's yeast IC ₅₀ (mM)	IC ₅₀ (mM)	IC ₅₀ (mM)	IC ₅₀ (mM)	IC ₅₀ (mM)	
Salacinol (13)	0.31 (9.6) Maltase ^e 0.32 (2.5) Sucrase ^e 0.47 (1.8) Isomaltase ^e	1.71	0.01	0.015	0.19	0.075	1.1×10^{-3}	NI	NI	_	_	$2.1^{\rm f,g} \sim 7.5^{\rm h}$	22,35,38, 39,46,55, 64,107
Kotalanol (14)	0.23 Maltase 0.18 Sucrase 1.8 Isomaltase	_	_	_	_	_	_	_	_	_	_	_	23
15		2.17	>5	>5	NI	NI	_	_	_	_	_	_	38,46,50
17	_	1.06	>5	>5	NI	NI	0.38	NI	3.4	_	_	$\begin{array}{l} 3.6^{\rm f} \\ \sim 7.5^{\rm h} \end{array}$	38,46,50, 64,107
Heteroatom analo	gues of salacinol												
Ghavamiol (25)	306 Maltase ⁱ 44 Sucrase ⁱ 136 Isomaltase ⁱ >315 Trehalase ⁱ	>2.5	>5	>5	NI	NI	_	_	_	_	_	~7.5 ^h	38,39,46, 50,107
26	_	>8	>5	>5	NI	0.4	_	_	_	_	_	_	38,46,50
Blintol (29)	_	0.72	>5	>5	0.49	>5	_	_	_	_	_	${\sim}7.5^{\rm h}$	38,44,46, 55,107
30	_	>9	>5	>5	NI	NI	_	_	_	_	_	$\sim 7.5^{\rm h}$	38,44,46,1
Five-membered rir	าช analogues												
37	_	>5	0.052	0.109	_	_	_	_	_	_	_	_	50
38	_	>30	>5	>5	_	_	_	_	_	_	_	_	50
42	_	_	_	_	NI	_	_	_	_	_	_	_	52
43	_	_	_	_	NI	_	_	_	_	_	_	_	52
65	_	_	_	_	_	_	NI	NI	NI	NI	0.853^{j}	NI	63
66	_	_	_	_	_	_	NI	NI	$2.47^{j,k}$	NI	0.716^{j}	NI	63
67 + 68	_	_	_	_	_	_	NI	NI	$1.87^{j,k}$	NI	NI	NI	63
69 + 70	_	_	_	_	_	_	NI	NI	NI	NI	1.29 ^{j,k}	NI	63
Six-membered rin	g analogues												
55	_	NI	_	_	NI	_	_	_	_	_	_	_	55,59
56	_	NI	_	_	_	_	_	_	_	_	_	_	59
57a, 58, 59a	_	NI	_	_	NI	_	_	_	_	_	_	_	46,59
57b	_	NI	_	_	NI	>5	1.41 ^{j,k}	NI	0.016 ^j	NI	NI	0.467 ^j	38,46,55, 59,63
59b	_	NI	_	_	NI	>5	_	_	_	_	_		38,46,59
71	_	_	_	_	_	_	NI	NI	NI	_	$0.85^{j,k}$	$1.34^{j,k}$	63
72	_	_	_	_	_	_	$1.32^{j,k}$	NI	NI	NI	NI	3.59 ^{j,k}	63
77–80	_	_	_	_	_	_	NI	NI	NI	NI	NI	NI	65
Seven-membered i	ring analogues												
73	_	_	_	_	_	_	NI	NI	NI	NI	NI	NI	63
74	_	_	_	_	_	_	NI	_	NI	_	NI	$1.83^{j,k}$	63
Chain-extended ar	nalogues												
94	_	_	_	_	NI	_	_	_	_	_	_	_	72
95	_	_	_	_	0.25	_	_	_	_	_	_	_	72
96	_	_	_	_	0.26	_	_	_	_	_	_	_	72
97	_	_	_	_	0.17	_	_	_	_	_	_	_	72
106	_	_	_		41	_	_		_		_	_	77

107	_	_	_	_	NI	_	_	_	_	_	_	_	77
108	_	_	_	_	26	_	_	_	_	_	_	_	77
109	_	_	_	_	NI	_	_	_	_	_	_	_	77
110	_	_	_	_	25	_	_	_	_	_	_	_	77
111	_	_	_	_	5	_	_	_	_	_	_	_	77
117	_	_	_	_	0.65	_	_	_	_	_	_	_	78
118	_	_	_	_	0.14	_	_	_	_	_	_	_	78
122	_	_	_	_	0.10	_	_	_	_	_	_	_	79
123	_	_	_	_	0.10	_	_	_	_	_	_	_	79
124	_	_	_	_	35	_	_	_	_	_	_	_	79
125	_	_	_	_	8	_	_	_	_	_	_	_	79
Deoxy-salacin	ol analogues												
128	>1320 Maltase ⁱ	_	_	_	_	_	_	_	_	_	_	_	82
120	780 Sucrase ⁱ												02
129	>1260 Maltase ⁱ	_	_	_	_	_	_	_	_	_	_	_	82
	>1260 Sucrase ⁱ												02
130	>1390 Maltase ⁱ	_	_	_	_	_	_	_	_	_	_	_	82
	>1390 Sucrase ⁱ												
Frame-shifted					20								0.0
138	_	_	_	_	20	_	_	_	_	_	_	_	89
139	_	_	_	_	53 NI	_	_	_	_	_	_	_	89
140	_	_	_	_		_	_	_	_	_	_	_	89
141	_	_	_	_	NI	_	_	_	_	_	_	_	89
De-O-sulfonat													
146	15.6 Maltase ⁱ	_	_	_	_	_	_	_	_	_	_	_	90
	3.7 Sucrase ⁱ												90
147	14.0 Maltase ⁱ	_	_	_	_	_	_	_	_	_	_	_	90
	3.5 Sucrase ⁱ												
Carboxylate a	nalogues												
148	_	_	_	_	21	_	_	_	_	_	_	0.3 ^h	91
149	_	_	_	_	NI	_	_	_	_	_	_	NI^h	91
156	_	_	_	_	10	_	_	_	_	_	_	_	95
157	_	_	_	_	NI	_	_	_	_	_	_	_	95
	•												
Phosphate der 158	ivatives				NI								26,96
159	_	_	_	_		_	_	_	_	_	_	_	
	_	_	_	_	NI	_	_	_	_	_	_	_	26,96
	ol-related analogues												
177–184	_	_	_	_	NI	_	_	_	_	_	_	_	99
211-219	_	_	_	_	6–75	_	_	_	_	_	_	_	106

NI: no inhibition.

^a β-Glucosidase from almond.

^b α-Galactosidase from green coffee beans.

^c β-Galactosidase from *Aspergillus oryzae*.

 $^{^{\}rm d}$ α -Mannosidase from jack beans.

^e Values in parentheses indicate IC₅₀ values.

f α -Mannosidase from almond. g 40% Inhibition.

^h Drosophila melanogaster Golgi mannosidase II.

 $^{^{}i}$ IC₅₀ values in μ M.

 $^{^{}j}$ K_{i} values in mM.

^k Preliminary determined with $K_{\rm m}$ and one $K'_{\rm m}$.

comparable to the placebo control group. 113 As it appears that the use of sulfonium salts as glycosidase inhibitors has also been exploited by Nature, the design of sulfonium-ion mimics could be a promising approach in the search for novel inhibitors. Various structure—activity studies of salacinol and related compounds have provided the key structural elements that appear to be necessary in the design of salacinol-based inhibitors. Such knowledge is essential if salacinol or its analogues are to be used in the treatment of type II diabetes as well as to gain further insights into their mode of action. Inhibition data for the compounds described in this review are listed in Table 3.

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